

Modelling plant-nematode interactions to understand plant tolerance

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MacBiosCore seminar 2023



Outline

- 1 Context
- 2 Plant model
- 3 Plant-Nematode interactions
- 4 Model calibration

Context

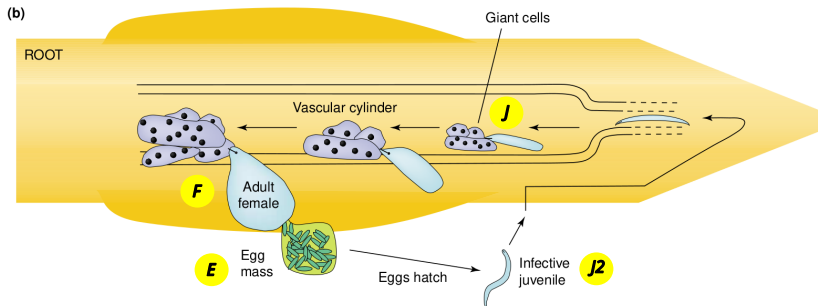
Root-knot nematodes

- microscopic endoparasites feeding on plant roots
- more than 5000 host species concerned
 - changes in root architecture
 - wilting of vegetative part, yield reduction
- 14% of global crop losses worldwide

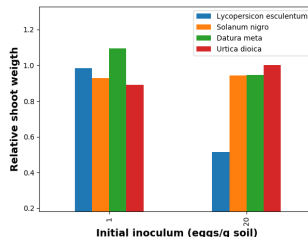
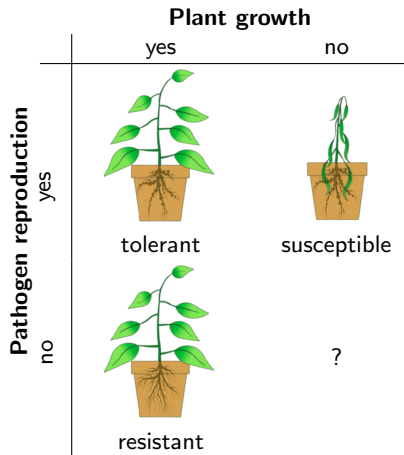


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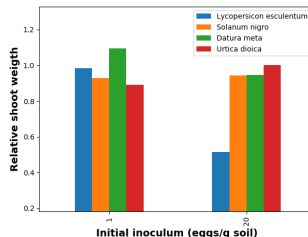
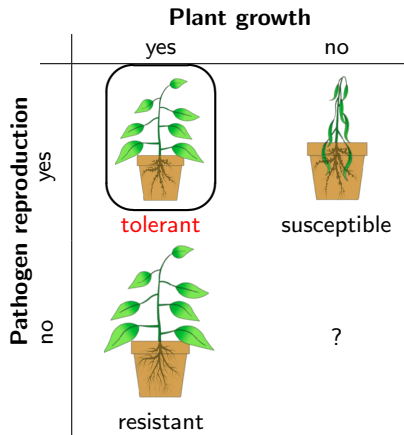


- **Strong variations** in the extent of damages within and among plant species



- Resistance is efficient but
 - ▶ few resistant genes identified
 - ▶ only for a few species
 - ▶ may favour the emergence of virulent nematodes
- Interest for tolerance
- Several possible mechanisms reported
 - ▶ high root growth rate
 - ▶ better water and nutrients uptake
 - ▶ delayed senescence ...

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Aim

Better understand the mechanisms behind (differential) plant susceptibility to nematodes

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Research question

Which are the **physiological mechanisms** behind **plant tolerance**?

Phenotypic variability

Aim

Better understand the mechanisms behind (differential) plant susceptibility to nematodes

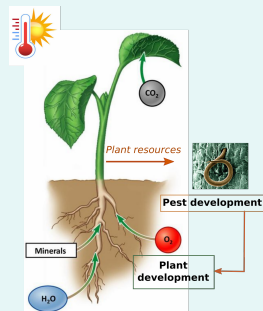
Research question

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Approach

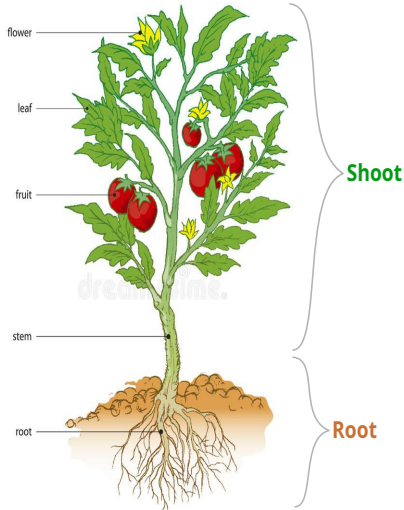
Coupled modeling of plant physiology and pest population dynamics

- better representation of pathosystem dynamics
 - interplay between plant and pest dynamics
- better prediction of plant growth rate and yield
- screening of interesting plant phenotypic traits



Plant model

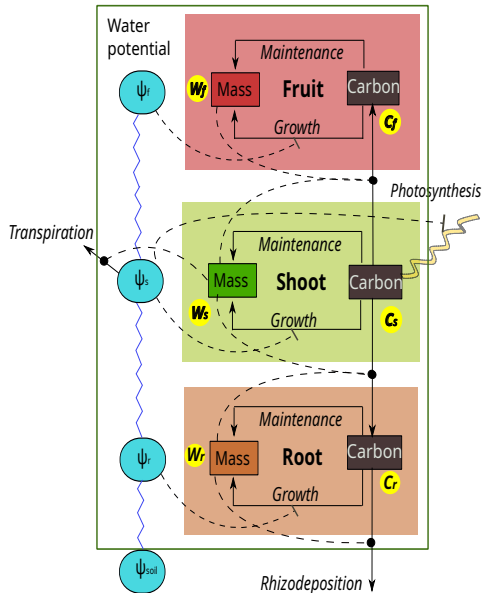
Plant description



The basic structure of plant (tomato).

VS

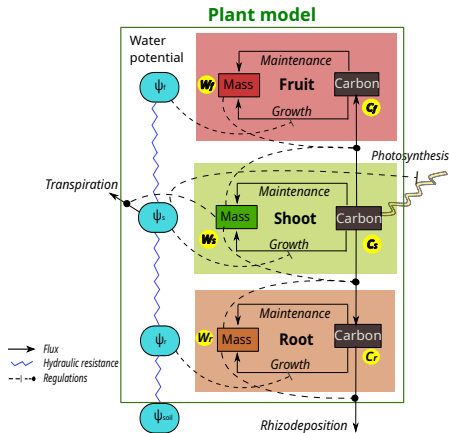
Plant model



Plant description

- 3 plant compartments:
 - ▶ shoots
 - ▶ roots
 - ▶ **fruits**
- 2 resources:
 - ▶ carbon (structural and free)
 - ▶ water
- Uptake, transport and allocation through plant compartments
 - ▶ growth and respiration processes
 - ▶ regulatory functions with respect to the water status of the plant

Thornley (1972), Dewar (1991)



$$\text{Shoot} \left\{ \begin{array}{l} \frac{dW_s}{dt} = \\ \frac{dC_s}{dt} = \end{array} \right.$$

$$\text{Root} \left\{ \begin{array}{l} \frac{dW_r}{dt} = \\ \frac{dC_r}{dt} = \end{array} \right.$$

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Mathematical equations

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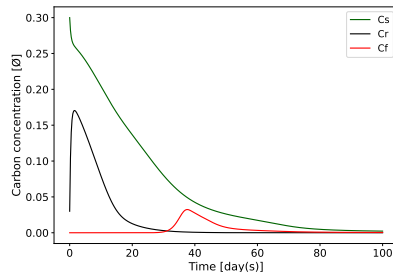
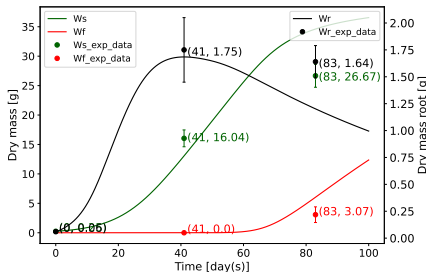
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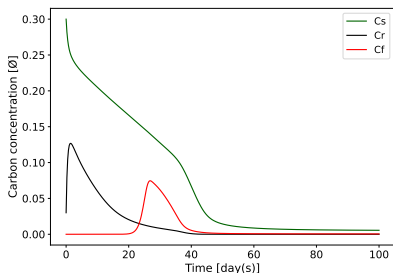
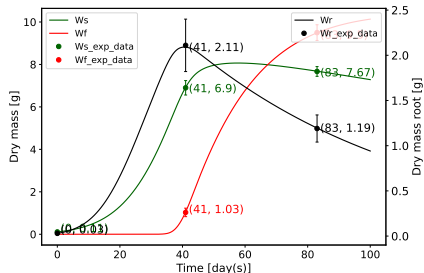
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Tomato plants



Pepper plants

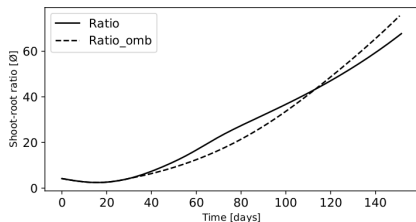


Test of plant model: environmental scenarios

The model is able to react correctly to a panel of environmental scenarios:

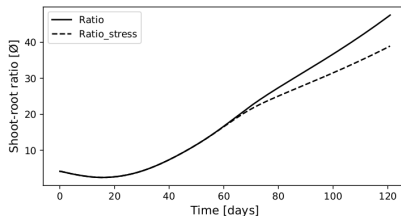
- allocation towards the most limiting resource

Shading



$$\sigma_c \rightarrow \sigma_c/2$$

Water stress



$$\Psi_{soil} \rightarrow -200MPa$$

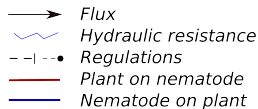
$$Ratio = \frac{W_s}{W_r}$$

Plant-Nematode interactions

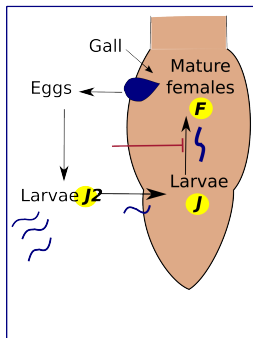
- Hijack of cell machinery amplifies **carbon sink in root galls** - *M.G.K. Jones. et al. 1981*
- **Increased hydraulic resistance** - *R. Dorhout, F.J. Gommers, 1991*
- Change in plant vascularization - *Bartlem et al. 2014*
- **Inhibition of photosynthesis** by water stress and/or interference between synthesis and translocation of growth hormones - *B. Loveys and A. F. Bird, 1994*
- Decrease of respiration intensity - *T. Mateille, 1994*
- Changes in glucose metabolism, nucleogenesis, proteogenesis... - *T. Mateille, 1994*

- Nematode growth rate and body size depend on food supply - *McLeod et al. 2001*
- **Host physiological stress affects egg production and sex ratio** - *Ferris et al. 1984, Snyder et al. 2006*
- **No effect of nitrogen fertilization** on nematodes development - *Spiegel et al. 1982*

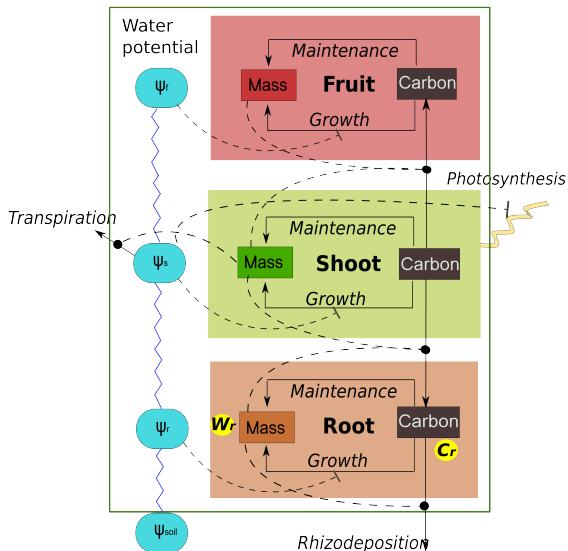
Plant-nematode model



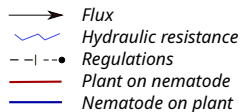
Pest model



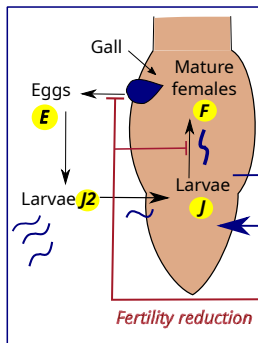
Plant model



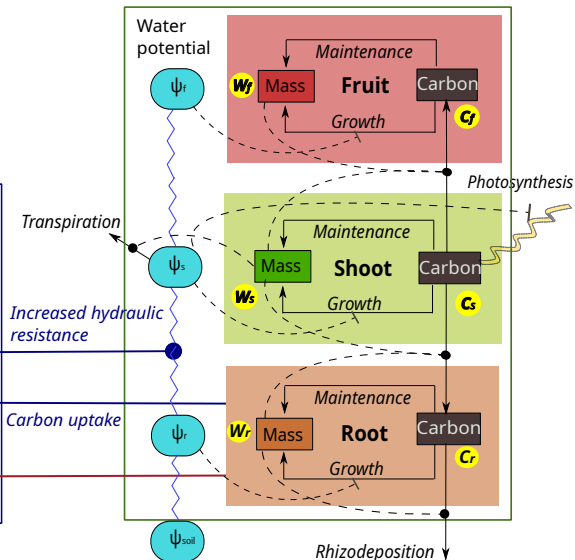
Plant-nematode model



Pest model



Plant model



RKN model:

$$\text{Free} \left\{ \begin{aligned} \frac{dE}{dt} &= \underbrace{r F}_{\text{Egg laying}} - \underbrace{h E}_{\text{Egg hatching}} - \underbrace{\mu_e E}_{\text{Mortality}} \\ \frac{dJ_2}{dt} &= \underbrace{h E}_{\text{Egg hatching}} - \underbrace{\beta J_2 W_r}_{\text{Larvae infection}} - \underbrace{\mu_{J_2} J_2}_{\text{Mortality}} \end{aligned} \right.$$

$$\text{Inside root} \left\{ \begin{aligned} \frac{dJ}{dt} &= \underbrace{\beta J_2 W_r}_{\text{RKN entry}} - \underbrace{\eta J}_{\text{Maturation}} - \underbrace{\mu_j J}_{\text{Mortality}} \\ \frac{dF}{dt} &= \underbrace{\eta J}_{\text{Maturation}} - \underbrace{\mu_F F}_{\text{Mortality}} \end{aligned} \right.$$

Plant roots:

$$\text{Root} \left\{ \begin{aligned} \frac{dW_r}{dt} &= \underbrace{k_r f(\psi_r) \frac{C_r}{K_r + C_r} W_r}_{\text{Growth}} - \underbrace{\gamma_r W_r}_{\text{Natural senescence}} - \underbrace{\gamma_r \left(\frac{K_m^n}{K_m^n + C_r^n} \right) W_r}_{\text{Additional drop due to carbon shortage}} \\ \frac{dC_r}{dt} &= \underbrace{\frac{1}{W_r} T_r}_{\text{Transport}} - \underbrace{(f_c + r_{g,r}) k_r f(\psi_r) \frac{C_r}{K_r + C_r}}_{\text{Growth}} - \underbrace{r_{m,r} \left(\frac{C_r^n}{K_m^n + C_r^n} \right)}_{\text{Maintenance respiration}} - \underbrace{c_{rh} \frac{1}{W_r^{(1-\beta)}} C_r}_{\text{Rhizodeposition}} - \underbrace{\frac{1}{W_r} \frac{dW_r}{dt} C_r}_{\text{dilution}} \end{aligned} \right.$$

RKN model:

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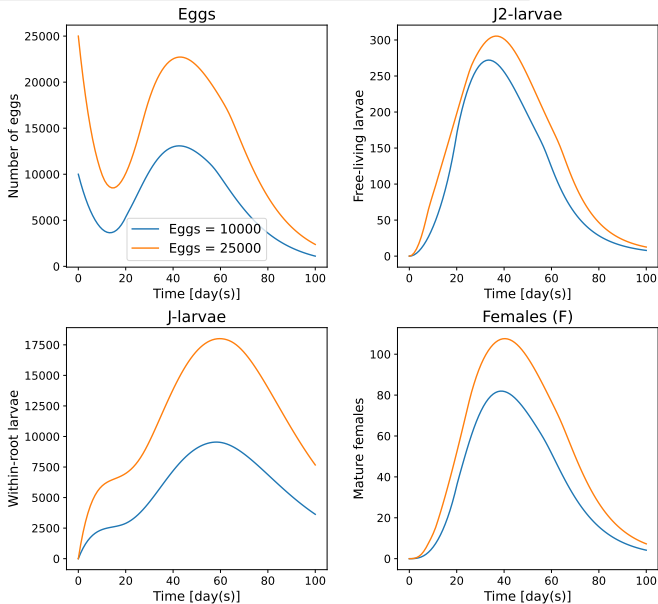
Plant roots:

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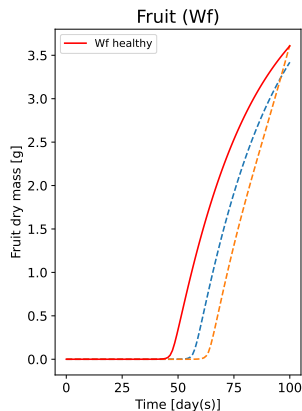
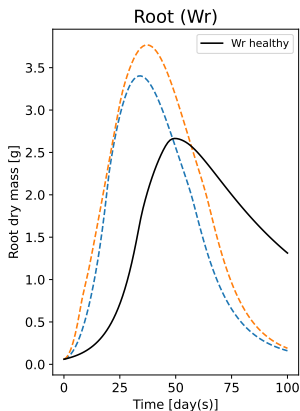
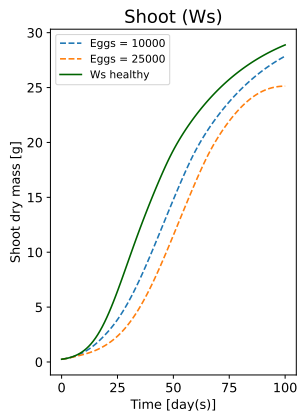
Plant → Nematodes effects

Nematodes → Plant effects

Preliminary Simulation plant-pest model



Preliminary Simulation plant-pest model



Model calibration

- Highly complex and non-linear model
- More than 30 parameters
- Few data available in literature

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- Dedicated experiments

- 2-steps calibration strategy:

- 1 Calibration of the plant model on data from healthy plants
 - ▶ data processing
 - ▶ global sensitivity analysis
 - ▶ numerical estimation of sensitive parameters
- 2 Calibration of the plant-nematode model on data from infected plants
 - ▶ global sensitivity analysis: nematode parameters
 - ▶ numerical estimation of sensitive parameters

Plant model Calibration

- Dedicated experiments
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Work in progress!

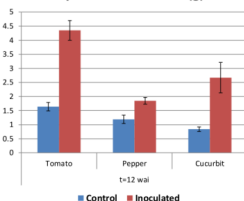


ArchiNem project (INRAE 2020-21)

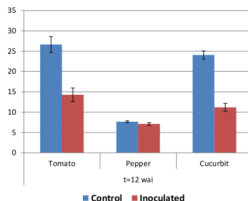
Experimental data available on **3 contrasted species** (tomato, pepper, cucurbit)

- Plant growth, physiology (healthy and infected)
 - Shoot, root, fruit dry mass (destructive): **3 time-points**
 - Leaf number: **12 time-points**
- 6 replicates per time-point

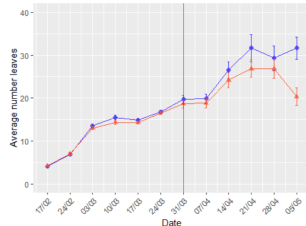
Dry mass of the roots (g)



Dry mass of the aerial parts (g)



Average number of leaves for tomato plants (cv. St Pierre)



Collaboration with C. Caporalino (INRAE)

Objective: increase the number of data points by exploiting non-destructive measurements (leaf number)

Allometric relation: shoot weight vs leaf number

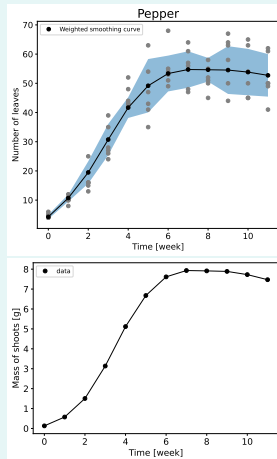
- Spline smoothing of leaf data
 - ▶ average dynamics
- Calibration of a general allometric relation (3 time-points x 3 species)

$$M = a_i L^b, \quad i = \text{species } 1, 2, 3$$

M = shoot dry mass, L = number of leaves



- Prediction of shoot dry mass from leaf data at 12 time-points



Multi-variate global sensitivity analysis

inputs : 26 parameters, reference value $\pm 30\%$

outputs : Shoot and root dry masses *dynamics* (vector)

method: R `multisensi` package (PCA- and ANOVA-based)

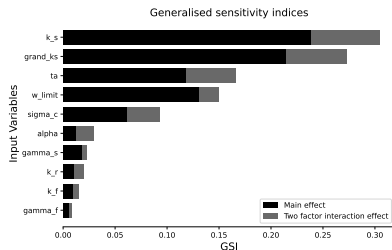
Multi-variate global sensitivity analysis

inputs : 26 parameters, reference value $\pm 30\%$

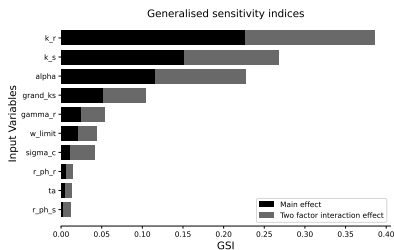
outputs : Shoot and root dry masses *dynamics* (vector)

method: R multisensi package (PCA- and ANOVA-based)

Shoot growth



Root growth



8 sensitive parameters (1 parameter fixed from literature data):

k_s shoot growth rate

k_r root growth rate

σ_c shoot carbon fixation rate

K_s shoot growth dynamics

t_a active C transport to fruit

γ_r root senescence rate

α vascular architecture

w_{limit} water stress sensitivity

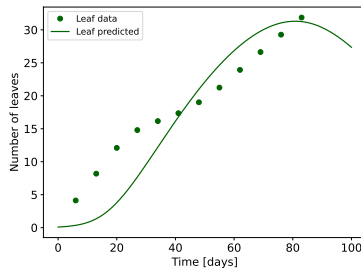
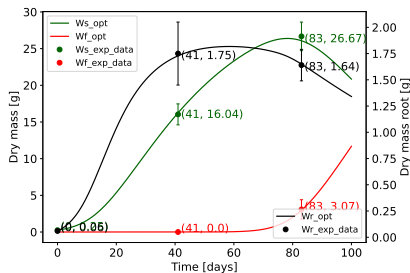
- 1 calibration per species considered
- Estimation of 7 (sensitive) parameters
- Weighted (π) least-square function for real and reconstructed mass data:

$$\underbrace{\frac{\pi}{N_i} \sum_{i=1}^{N_i} \left(\frac{M_{sol}^i - M_{data}^i}{std^i} \right)^2}_{\text{Dry mass data}} + \underbrace{\frac{(1-\pi)}{N_j} \sum_{j=1}^{N_j} \left(\frac{M_{sol}^j - M_{pred}^j}{M_{pred}^j} \right)^2}_{\text{Reconstructed data from leaf number}}$$

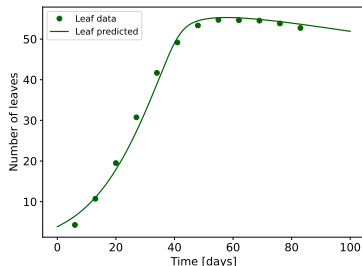
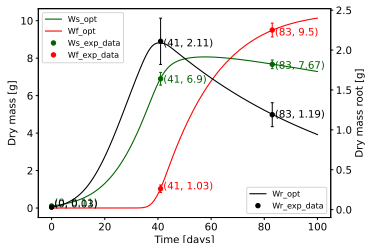
2-step estimation strategy:

- 1 Global search (differential evolution algorithm)
- 2 Local search around solution of step 1 (least square)

Tomato plants



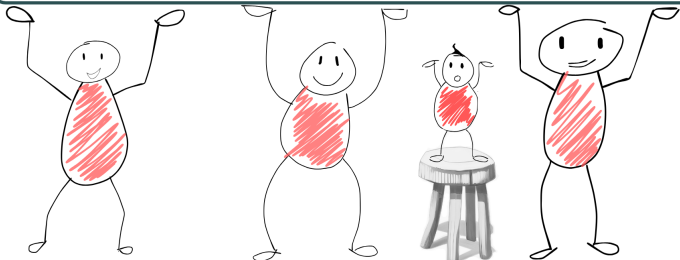
Pepper plants



- Calibration of RKN-related parameters on data from infected plants
- Identification of key physiological and architectural traits underlying **plant tolerance** to RKN infestation
- **Multi-seasonal**² effect of plant tolerance: **crop rotations** (tolerant, susceptible and resistant plants), management practices, effect of environmental conditions...

²Nilusmas et al., *Evolutionary Applications*, 2020

THANK YOU FOR YOUR ATTENTION!



Collaborators:

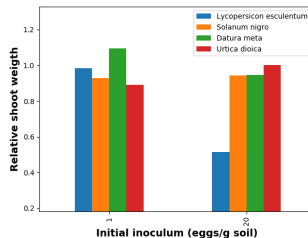
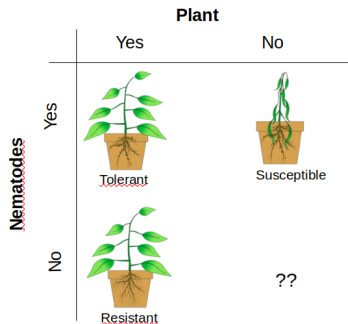
INRAE colleagues:

- C. Caporalino
- L. Pagès
- C. Doussan

Master students

- T. Brenière
- N. Juazion-Graverolle
- C. Bourgade
- E. Ceci
- A. Canaud

- **Strong variations** in the extent of damages within and among plant species



- Resistance is efficient but
 - ▶ few resistant genes identified
 - ▶ only for a few species
 - ▶ may favour the emergence of virulent nematodes
- Interest for tolerance
- Several possible mechanisms reported
 - ▶ high root growth rate
 - ▶ better water and nutrients uptake
 - ▶ delayed senescence ...

Aim

Better understand the mechanisms behind (differential) plant susceptibility to nematodes

Water transport

$$R_{sr} = r_{sr} (W_r + G)^{-\alpha_r} \quad (1)$$

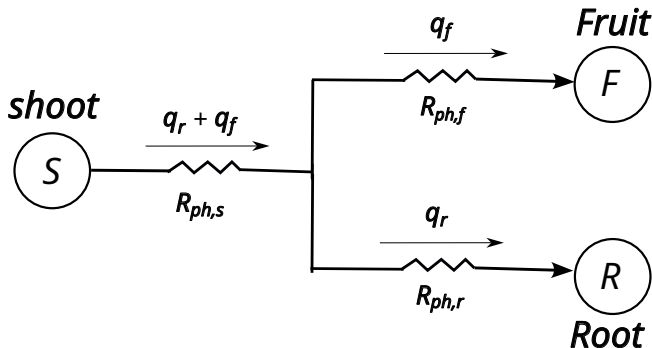
$$R_{xy} = \frac{r_{xy}}{W_r^{\alpha_r}} + \frac{r_{xy}}{W_s^{\alpha_s}} \quad (2)$$

Carbon transport

$$R_{ph} = \frac{r_{ph}}{W_s^{\alpha_s}} + \frac{r_{ph}}{(W_r + G)^{\alpha_r}} \quad (3)$$

Carbon transport (T)

The carbon flow³ T in phloem vessels $T = q C_s$, with q the volume flow rate,



Therefore,

$$\begin{cases} (q_r + q_f)R_{ph,s} + q_r R_{ph,r} = (C_s - C_r) \\ (q_r + q_f)R_{ph,s} + q_f R_{ph,f} = (C_s - C_f) \end{cases}$$

$$\begin{cases} T_r = q_r C_s \\ T_f = \left(\frac{W_s^n}{I^n + W_s^n} \right) q_f C_s \end{cases}$$

³Minchin et al., *Journal of Experimental Botany*, 1993

Male proportion

$$\theta(C_r) = \theta_{\max} \frac{K_{\theta}}{C_r + K_{\theta}} \quad (4)$$

Reproduction rate

$$r(C_r) = r_{\min} + \frac{(r_{\max} - r_{\min}) C_r}{C_r + K_{repro}} \quad (5)$$

Establishment of the feeding site

$$\Omega(C_r) = \frac{Wcr^{\kappa}}{Wcr^{\kappa} + K_{\Omega}^{\kappa}} \quad (6)$$

Goal: Identifying the most influential parameters

inputs : 26 parameters, **outputs** : dry masses along time (vector)

Method

- 1 fractional factorial design to explore parameter space
- 2 PCA to reduce output and capture its variability
- 3 ANOVA to compute sensitivity indices (SI)

$$SI_{..} = \frac{SS_{..}}{TSS}, \quad GSI = \sum_{k=1}^{components} SI_k \times inertia_k$$

SS = sum of squares, TSS = total sum of squares

Parameter set

TABLE 1 – Standard values for sensitivity analysis

Parameters	Description	Value	Interval
σ_C	Specific carbon fixation rate by the aerial part	0.4	$\pm 30\%$
σ_W	Specific transpiration rate of aerial part	1.2	$\pm 30\%$
$r_{ph,s}, r_{ph,r}, r_{ph,f}$	Xylem resistance coefficients to water flow	0.5, 0.5, 0.005	$\pm 30\%$
$r_{xy,s}, r_{xy,r}, r_{xy,f}$	Phloem resistance coefficients to sap flow	5, 5, 0.1	$\pm 30\%$
r_{sr}	Coefficient of resistance to water absorption by the roots	1	$\pm 30\%$
k_s, k_r, k_f	Specific growth rates	0.14, 0.12, 0.6	$\pm 30\%$
K_s, K_r, K_f	Half-saturation constants related to growth rate	0.1, 0.05, 0.01	$\pm 30\%$
$\gamma_s, \gamma_r, \gamma_f$	Mortality and leaf fall coefficients	0.1, 0.01, 0.001	$\pm 30\%$
n, n_c, n_p, n_{hill}	Hill coefficients for transition, growth, photosynthesis and water regulation	10, 10, 10, 10	$\pm 30\%$
W_{limit}	Transition coefficient from vegetative to reproductive phase	15	$\pm 30\%$
K_c, K_p	Half-saturation constants related to the effect of water on growth and photosynthesis	-1400, -1600	3 fixed values
$\alpha_s, \alpha_r, \alpha_f$	Allometric coefficients	2/3, 2/3, 2/3	$\pm 30\%$
$r_{s,m}, r_{r,m}, r_{f,m}$	Maintenance respiration coefficients for aerial, root and fruit compartments	0.001, 0.001, 0.001	$\pm 30\%$
$r_{s,g}, r_{r,g}, r_{f,g}$	Growth respiration coefficients of aerial, root and fruit compartments	0.01, 0.01, 0.01	$\pm 30\%$
c_{rh}	Rhizodeposition coefficient	0.1	$\pm 30\%$
ta	concentration rate of carbon related to active transport	0.5	$\pm 30\%$

- 26 parameters
- 3 levels for each parameter (reference value and $\pm 30\%$ of reference value)
- some reference values from literature